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Septoria malagutii as an endophytic fungus of *Achillea millefolium* from Iran

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From May until September 2015, a survey was conducted to collect endophytic fungi from healthy medicinal plants in the natural ecosystem of Golestan province. Endophytic fungi were isolated from *Achillea millefolium* plants. Different samples were provided from roots, leaves and stems of the healthy and mature plants. The plants were rinsed gently under running water and samples were cut into the 0.5–1 cm pieces. The surface sterilization was performed by sodium hypochlorite (NaOCl) and 75% ethanol. The surface sterilized samples were placed on PDA plates supplemented with 50 mg/L tetracycline to suppress the bacterial growth and incubated at $28 \pm 2^\circ\text{C}$ up to 14 days. Morphological identification was performed using fungal identification keys (Cline & Rossman 2006; Sutton 1980; Priest 2006, Verkley et al. 2013). Subsequently, the nucleic acid was extracted using the cetyl trimethyl ammonium bromide (CTAB) method (Doyle 1990). The strains were sequenced partially for four genomic regions including internal transcribed spacer regions (ITS) of nrDNA, 28S nrDNA gene (LSU), translation elongation factor 1- α (*tefl- α*) and β -*tubulin* (*tub2*). All

the sequences were deposited in NCBI's GenBank Database.

A basic alignment of the obtained sequence data was done using MAFFT v. 7 and Mr Modeltest 2.3 was used to determine the best substitution models for each locus. Bayesian analyses were performed on the concatenated loci using MrBayes v. 3.2.1 (Huelsenbeck & Ronquist 2001).

Based on the four-regions tree, the examined strain was grouped in the same clade with *S. malagutii* (CBS 106.80) and identified as *Septoria malagutii*.

The genus *Septoria*, is one of the largest genera of plant pathogens, causing a range of disease symptoms including leaf and fruit spots (Verkley et al 2013). However, the endophytic species of this genus had not yet been reported. This is the first report of *S. malagutii* as an endophytic fungus from Iran (Ershad 2009).

Septoria malagutii E.T. Cline, Mycotaxon 98: 132 (2006).

Septoria malagutii have been isolated as an endophyte from *A. millefolium* in the Golestan province with (36°36'10"N 54°29'55"E) geographical coordinates. Morphology on PDA after 15 days of culture at 23°C: colony 22 mm in diameter, aerial mycelium white, underlying color grey to black, surface floccose, slightly raised and reverse faintly ringed. Conidiomata pycnidial, slightly paler, epigenous, scattered, globose to subglobose, solitary, smooth, 97 μm diameter, formed mostly on the agar surface, ostiole was not observed, immature pycnidia black, 43–70 μm diameter. Conidiophores were not observed. Conidiogenous cells were ampulliform, rarely doliform, discrete, determinate, hyaline, 4–10 × 2–7 μm. Conidia holoblastic, hyaline, filiform, strongly curved, occasionally straight, sharply pointed at both ends, 4–6 septate, sometimes slightly rounded at base, 60–125 × 1.7–2 μm diameter. The fungal isolate was deposited in the Iranian Fungal Culture Collection (IRAN C) of the Iranian Research Institute of Plant Protection, Tehran, Iran with voucher code: IRAN 3261C. Accession numbers in the NCBI's GenBank Database: (ITS: MH259175, LSU: MH255545, *tefl- α* :MK952153), *tub2*:MK952152).

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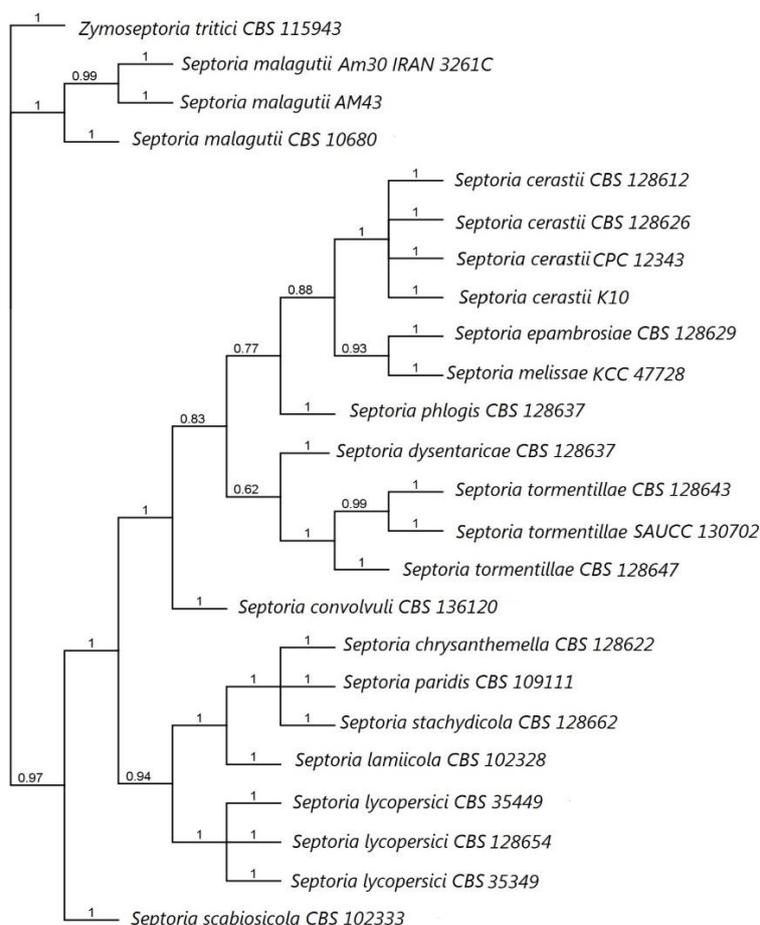


Fig. 1. Bayesian tree from a concatenated of ITS, LSU, *tefl* and *tub2* data sets. Numbers on the branches are Bayesian posterior probabilities (PP). The tree was rooted with *Zymoseptoria tritici* as outgroup taxa. IRAN 3261C is related to the *S. malagutii* in this study.

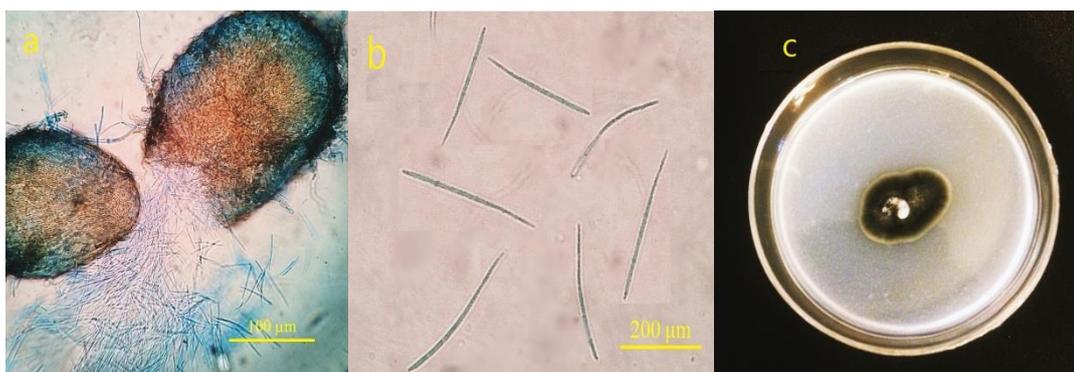


Fig. 2. *Septoria malagutii*: a. pycnidia, b. conidia, c. Colony on PDA (potato dextrose agar) after 14 days of culture.

REFERENCES

- Cline ET, Rossman AY. 2006. *Septoria malagutii* sp. nov., cause of angular leaf spot of potato. *Mycotaxon*. 98:125-135.
- Doyle JJ, Doyle JL. 1990. Isolation of plant DNA from fresh tissue. *Focus*. 12:13-15.
- Ershad DJ. 2009. *Fungi of Iran*. Agricultural research, Education & Extension Organization (AREEO), Ministry of Agriculture, Tehran, Iran.
- Huelsenbeck, J.P. & Ronquist, F. 2001. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 17: 754-755.
- O'Donnell K, Cigelnik E. 1997. Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus *Fusarium* are nonorthologous. *Molecular Phylogenetics and Evolution* 7:103-107.
- Priest MJ. 2006. *Fungi of Australia - Septoria*. CSIRO Publishing, Melbourne, Australia.

- Rehner SA, Buckley EA. 2005. Beauveria phylogeny inferred from nuclear ITS and EF1- α sequences: evidence for cryptic diversification and links to Cordyceps teleomorphs. *Mycologia* 97: 84-98.
- Sutton BC. 1980. The coelomycetes: fungi imperfecti with pycnidia, acervuli, and stromata. *Commonw. Mycol. Inst. Kew, Surrey, England*.
- Verkley GJM, Quaedvlieg W, Shin H-D, Crous PW. 2013. A new approach to species delimitation in *Septoria*. *Studies in Mycology* 75:213–305.
- Vilgalys R, Hester M. 1990. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* 172: 4239-4246 DOI: 10.1128/jb.172.8.4238-4246.1990.
- White TJ, Bruns T, Lee S, Taylor JW. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *PCR protocols: a guide to methods and applications*. (MA Innis, DH Gelfand, JJ Sninsky & White TJ, eds). 315-322.: Academic Press, New York, USA.